

We would like to thank Dr. JEROME VINOGRAD for his helpful advice and discussion.

This work was supported in part by a research grant, E-536 (C-4) from the National Institutes of Health, U.S. Public Health Service, and contract AT (11-1)-34, Project 8 of the Division of Biology and Medicine, U.S. Atomic Energy Commission.

*Botany Department, University of California,  
Los Angeles, Calif. (U.S.A.)*

ALBERT SIEGEL  
WILLIAM HUDSON

<sup>1</sup> M. MESELSOHN, F. W. STAHL AND J. VINOGRAD, *Proc. Natl. Acad. Sci. U.S.A.*, 43 (1957) 581.

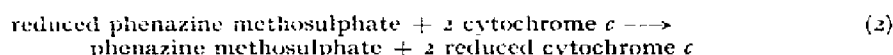
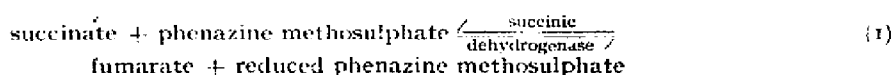
<sup>2</sup> J. S. JOHNSON, K. A. KRAUS AND T. E. YOUNG, *J. Am. Chem. Soc.*, 76 (1954) 1436.

<sup>3</sup> A. SIEGEL AND S. G. WILDMAN, *Phytopathology*, 44 (1954) 277.

Received November 10th, 1958

### The microestimation of succinate and the extinction coefficient of cytochrome *c*

Phenazine methosulphate has been shown to be a very good electron acceptor in the oxidation of succinate by succinic dehydrogenase<sup>1</sup>, whereas cytochrome *c* is an extremely poor acceptor in this system. However, cytochrome *c* is reduced rapidly in a non-enzymic reaction with reduced phenazine methosulphate<sup>2</sup>. Hence the oxidation of succinate by succinic dehydrogenase can be coupled to the reduction of cytochrome *c* in the presence of trace amounts of phenazine methosulphate to give an extremely sensitive method for the estimation of succinate.



The optimal conditions for this estimation have not been explored fully, but the following conditions have been found satisfactory. Spectrophotometer cells are filled with the following reagents to a total volume of 2.95 ml; phosphate, pH 7.6, 150  $\mu$ moles; cytochrome *c*, 0.125  $\mu$ mole; succinate, 0–0.05  $\mu$ mole; succinic dehydrogenase prepared according to the method of SINGER *et al.*<sup>1</sup>, 0.2 mg. The cells are incubated at 25° and the reaction begun by the addition of 0.05 ml of 0.01 % phenazine methosulphate (freshly diluted from a 1 % stock solution).

A blank without any succinate but containing all the other reagents must be run, since a red colour develops on the incubation of phenazine methosulphate with most proteins, due to a complex with sulphhydryl groups<sup>2</sup>. The absorbance at 550 m $\mu$  is recorded until the difference between sample and blank becomes constant. Fig. 1 shows the results of a number of estimations carried out in this way. It will be noted that the amount of cytochrome *c* reduced is the same in the presence or absence of oxygen. As reduced phenazine methosulphate is very auto-oxidizable (in fact its reaction with O<sub>2</sub> is the basis of the usual assay for succinic dehydrogenase<sup>1</sup>) it is evident that the rate of reaction of reduced phenazine methosulphate with cytochrome *c* must be very much greater than that of its re-oxidation with O<sub>2</sub>. As cyto-

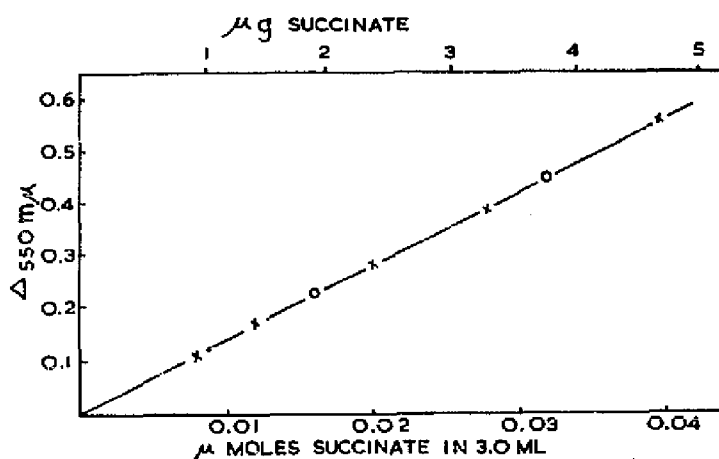


Fig. 1. The increase in absorbance at 550  $m\mu$  due to reduction of cytochrome *c* by succinate. The absorbancies plotted are after blank correction. Conditions, as in text. Enzyme used, 0.5 mg 40% pure succinic dehydrogenase. Succinate, neutralized Hopkins & Williams succinic acid, analytical grade. X aerobic, O anaerobic.

chrome *c* is a one-electron acceptor, and as phenazine compounds are reduced in two distinct one-electron stages<sup>3,4</sup>, it is not unreasonable to assume that the rapid reaction with cytochrome *c* is with the semiquinoid form of phenazine methosulphate, which in turn is probably formed from the semiquinoid flavin form of succinic dehydrogenase.

This coupled reaction of succinate oxidation and cytochrome *c* reduction may also be used to calculate the extinction coefficients of cytochrome *c*. There is considerable variation in the published values for the oxidized and reduced forms of cytochrome *c*, which have been based on calculations from dry weight or iron content. The reaction with succinate offers standardization against a stable compound of known purity and concentration, and therefore should be considerably more accurate. From the slope of Fig. 1 it can be calculated that  $\epsilon_{550}$  (reduced-oxidized) for cytochrome *c* is  $2.10 \cdot 10^4$   $\text{cm}^2/\text{mmole}$ . The highest value previously reported<sup>5</sup> is  $1.89 \cdot 10^4$ . The cytochrome *c* used in this work was prepared according to the method of KEILIN AND HARTREE<sup>6</sup> and further purified by fractionation on a column of calcium phosphate gel suspended on cellulose. Brown impurities were eluted with 0.1 *M* phosphate, pH 7.6 and the cytochrome *c* eluted with the same buffer + 5% (w/v)  $(\text{NH}_4)_2\text{SO}_4$ , leaving behind considerable amounts of greenish impurities. The ratio of absorbance at 550  $m\mu$  (reduced with  $\text{Na}_2\text{S}_2\text{O}_4$ ) to that at 280  $m\mu$  (oxidized with  $\text{K}_3\text{Fe}(\text{CN})_6$ ) was 1.30. The molar extinction coefficient of the oxidized form at 550  $m\mu$  was found to be  $0.89 \cdot 10^4$   $\text{cm}^2/\text{mmole}$  and of the reduced form,  $2.99 \cdot 10^4$   $\text{cm}^2/\text{mmole}$ .

Department of Biochemistry, University of Sheffield (Great Britain) VINCENT MASSEY

<sup>1</sup> T. P. SINGER, E. B. KEARNEY AND P. BERNATH, *J. Biol. Chem.* 223 (1956) 599.

<sup>2</sup> V. MASSEY, unpublished.

<sup>3</sup> L. MICHELIS, *Chem. Reviews*, 16 (1935) 243.

<sup>4</sup> H. McILWAIN, *J. Chem. Soc.*, (1937) 1704.

<sup>5</sup> E. MARGOLASH, *Biochem. J.*, 56 (1954) 535.

<sup>6</sup> D. KEILIN AND E. F. HARTREE, *Biochem. J.*, 39 (1945) 289.

Received November 20th, 1958